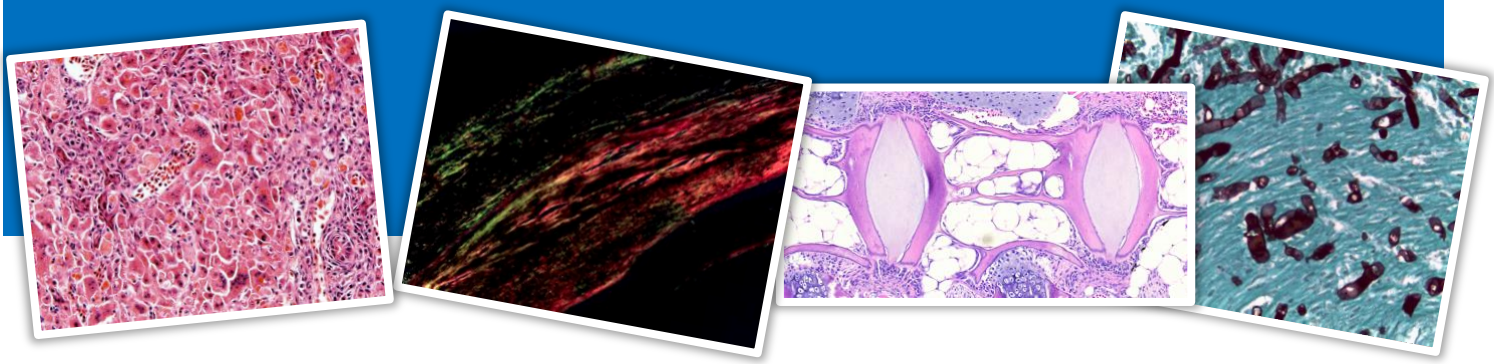
INSTRUCTION FOR HISTOPATHOLOGY SAMPLING



Objective

Describe the correct sampling of tissues for processing and subsequent histological diagnosis.

Materials and supplies: Scalpel handle, Scalpel blades, Scissors, Anatomic and surgical tweezers, Fixing solution: 10% buffered formalin, Container and Indelible marker pen.

Development of the process

a) Select the culture units (cages or tanks) and prepare a smooth surface for sampling (Image 1).

b) Label the containers with the fixing solution (10% buffered formalin) (Image 2). 10 parts of solution per 1 part of tissue are required (Image 3).

c) Samples should be obtained only from fresh tissue. Those samples that show the minimum degree of autolysis **DO NOT HAVE ANY DIAGNOSTIC VALUE**.

d) Scalpel cuts must be "clean", avoiding tearing the tissue. It is not recommended to use a rat tooth tweezers (Image 4).

e) **Sample size:** 0.5 cm thick (Image 5).

f) Fish less than 4 cm in length, introduce them completely in the fixative, making an incision along the abdominal cavity without damaging the digestive tract.

g) **Pyloric caeca:** make a longitudinal cut and take the anterior portion to obtain pancreatic tissue.

h) **Kidney:** Take preferably medium kidney.

i) **Gills:** lift the operculum, select first branchial arch and cut including a piece no larger than 0.5 cm. **Injury:** the sample must include normal and damaged tissue.

j) **Eyes:** the sample must include the complete eyeball.

k) Keep in a cool place between 1-20 ° C. Use gel pack if necessary. Do not expose to direct sunlight. Do not freeze.

Image 1: Materials for sampling

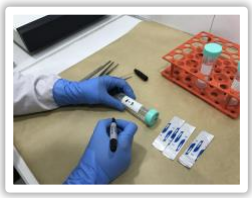


Image 2: Labeling of samples

Image 3: Fixing solution (10% bufferered formalin. 1:10 (tissue:solution)

Image 4: Clean tissue cutting

Image 5: Sample size, 0.5 cm thick

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